



PLANT-PARASITIC NEMATODES IN POME & STONE FRUIT ORCHARDS BIOLOGY & MANAGEMENT IN WASHINGTON

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A diversity of microscopic round worms called nematodes inhabit orchard soils. Many of the nematodes in orchard soils contribute to soil health functions. Fungal feeding and bacterial feeding, predatory and omnivorous nematodes provide a number of services including nitrogen mineralization and plant pathogen suppression.

However, plant-parasitic nematodes feed on plants; damaging roots and creating wounds which may serve as infection points for plant pathogenic fungi, thereby directly transmitting certain pathogens, such as viruses, to the host plant. Plant-parasitic nematodes have a needle-like stylet in their mouthparts which they use to feed on plant cells. Nematodes can either be migratory or sedentary. Migratory nematodes move through roots as they feed, disrupting plant cells or feeding entirely on the outer surface. Sedentary nematodes manipulate normal cell development and create a feeding site. All of these lifestyles and behaviors interrupt the plant's ability to transport water and nutrients. Nematode feeding can also directly kill plant cells, root hairs, and even whole roots.

Here we will describe a few key plant-parasitic nematodes that can cause damage in the orchard.

Lesion Nematode

(*Pratylenchus penetrans* and *P. vulnus*)

Lifecycle and Damage

Lesion nematode (Figure 1) is considered a migratory endo- and ectoparasite. This means that it can be found living and reproducing in the soil and feeding both on the outside and the inside the root. Feeding of the nematode kills root cells, and when the nematodes move through the root they damage and disrupt root tissue inhibiting water and nutrient movement into the plant. Feeding damage also provides access channels for fungi, bacteria, and other organisms. Life cycles vary from 45 to 65 days (Smiley 2010).

Symptoms

Young trees, especially apples, infected with lesion nematodes exhibit poor growth. Yields of infected trees may be significantly lower than non-infected trees. Infected roots generally have discolored and necrotic (black) areas where nematode feeding has caused cell death and where additional pathogens have likely invaded. Severely affected root systems may lack feeder roots. However, these symptoms may be difficult to distinguish from those caused by other pathogens on tree roots.

Thresholds

Lesion nematode thresholds for Washington orchards are not well established. However, based on experience in other states, pre-establishment action thresholds per soil sample of 500 cc (approximately one pint) are: (A) 0–10 crop should not be affected; (B) 20–70 may cause crop damage; (C) 80+ will likely cause damage.



Figure 1. Lesion nematode. Photo credit Howard Ferris, UC Davis.

Dagger Nematodes

(*Xiphinema americanum*, *X. rivesi*)

Dagger nematode is an economic concern in orchards because it can vector viruses including tomato ringspot virus (TRSV), the cause of apple union necrosis and decline, the cherry rasp leaf virus (CRLV) which causes flat apple, and peach rosette mosaic virus (PRMV).

Lifecycle and Damage

Dagger nematode is a migratory ectoparasite which means it attacks roots from the outside. It is a relatively large nematode (1.3 to 2.2 mm) (Akinbade et al. 2014). Dagger nematode has a long, hollow, needle-like stylet called an odontostyle which it uses to feed on cells at root tips. Feeding can cause root tip cells to enlarge, disrupting function and resulting in roots that have a knobby, stunted appearance, with swellings at the tips. While its life cycle has not been experimentally confirmed, researchers suggest that it may be up to one year in duration with individuals living three to five years. Dagger nematode can harbor virus particles in the esophageal lumen near the stylet extension and in the glandular region of the esophagus. As the nematode feeds, virus particles can be transmitted to the plant within 24 hours of exposure (Ferris 2017). Since the stylet, the stylet extension, and the lining of the esophageal lumen are shed when the nematode molts, virus particles are also shed as the nematode passes from one life stage to another. However, virus particles are rapidly reacquired from the plant on which the nematode is feeding.

Symptoms

Above-ground symptoms of nematode damage can be hard to detect and easily confused with other plant stresses resulting from water or nutrient deficiencies. The most commonly recognized symptoms that can be attributed to this pest are those that result from transmission of the associated viruses.

Plant symptoms resulting from viral infection commonly include: chlorotic mottling and defoliation; bright yellow discoloration of foliage; yellowing along main leaf veins; spots, blotches, and crinkling of leaves; and stunting. As with most nematodes the presence and abundance may be most readily determined by soil sampling.

Threshold

Nematode damage thresholds in Washington are not well established. However, based on experience in other states pre-establishment management thresholds per 500 cc soil sample (approximately one pint) are: (A) 0 (zero) no crop damage, (B) 1–10 may cause crop damage, (C) 20+ likely to damage. The threshold is so low (one or greater) due to the potential to vector viruses.

Ring Nematode

(*Mesocriconema* sp.; listed as *Criconemella* and *Criconematodes* in earlier literature.)

Ring nematode is common to soft fruit orchard soils, and the bacterial canker pathogen, *Pseudomonas syringae*, in concert with freeze damage, has been associated with the phenomenon termed “peach tree short life” (Cao et al. 2006, Ritchie and Clayton 1981). Root feeding in pome and stone fruit can result in loss of up to 85% of small roots causing significant water and nutrient stress (Westerdahl and Duncan 2015). As higher populations of *Mesocriconema* are known to occur in sandy soils with large pore spaces, this nematode deserves greater scrutiny when examining nematode populations in coarse-textured Washington orchard soils. However, it is not currently known to be an economic concern in Washington.

Lifecycle and Damage

Ring nematodes feed on roots from the outside with a long spear-like stylet. They feed primarily on fine roots killing them, and stunting growth of lateral roots.

Symptoms

Trees may show water and nutrient stress due to small, damaged root systems. Trees with no noticeable vigor issues directly from nematode feeding may develop bacterial canker particularly under conditions where cold damage has been experienced.

Threshold

Ring nematode damage thresholds in Washington are not established.

Pin Nematode

(*Paratylenchus spp.*)

Pin nematode is a minor pest in orchards.

Lifecycle and Damage

Pin nematode is a small nematode which feeds either singly or in groups on the outside of plant roots (Figure 2). Feeding causes small lesions and death of lateral roots. In general, it is found randomly distributed in the orchard across tree rows and alleyways (Howland et al. 2014).

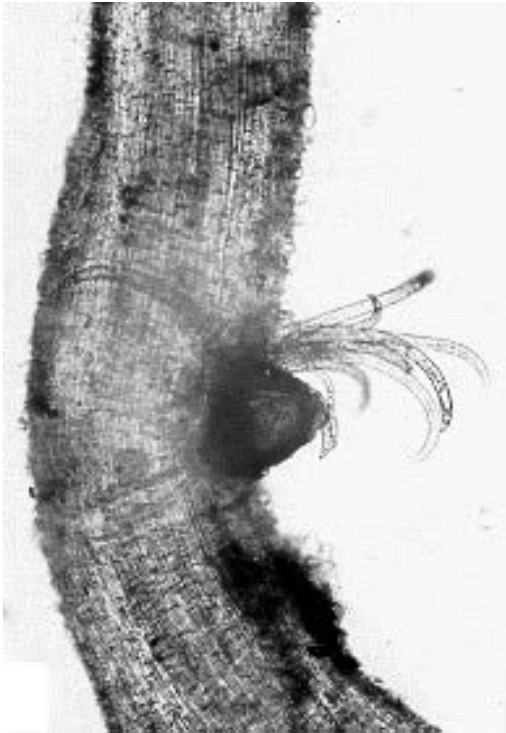


Figure 2. *Paratylenchus neoamblycephalus* feeding. Photo credit Lownsbery, 1975.

Symptoms

Small root lesions may be noticeable. General plant decline may be difficult to diagnose.

Threshold

Damage thresholds for pin nematode are not established for orchards.

Root-knot Nematode

(*Meloidogyne hapla*, *Meloidogyne chitiwood*)

Most root-knot nematode species have a wide host range affecting tree fruit as well as most small fruits, vegetables, and grains (Figure 3). Known for the gall formation it induces on plant roots, root knot nematode can cause plant nutrient and water stress by disrupting plant root function.



Figure 3. Root-knot nematode galls (strawberry roots). Photo credit M. Mazzola USDA-ARS.

Lifecycle and Damage

Root-knot nematode is a sedentary endoparasite. This means that while juveniles live in the soil outside the roots, adult females live and feed inside host roots. The second stage juvenile invades the root and establishes the feeding site. When root-knot nematodes insert their needle-like stylet into plant cells they inject proteins

that make the cells grow abnormally. Instead of growing and dividing, the cells create additional nuclei and become larger. This ‘giant-cell’ produces copious proteins which the nematodes then ingest.

Abnormal root development inhibits root function, resulting in a diminished capacity of roots to efficiently uptake water and nutrients (Mitkowski and Abawi 2003). Adult root-knot nematodes are generally concentrated in the irrigated root zone in vineyards and are likely to be concentrated in the irrigated root zone in Washington orchards (Howland et al. 2014). Most root-knot nematode species have a very wide host range.

Symptoms

Root-knot nematode feeding can cause large galls on infected plant roots. Plants may show signs of water stress or nutrient deficiency due to compromised root systems.

Threshold

Root-knot nematode damage thresholds are not established for orchards.

Field Evaluation

In order to determine nematode numbers in the orchard, sample soil in the root zone, either at the tree dripline, edge of the weed strip, or near drip emitter lines. At least 15 cores per five-acre block should be taken to a depth of at least 10 to 12 inches. In orchards with full size (non-dwarfing) trees a second sample from 12 to 24 inches is advisable. Plant feeding nematodes are in the highest concentration in the root zone. Mix the 15 or more subsamples in a clean bucket and remove a sample of one pint and place it in a plastic bag to send to a nematode testing lab. Remember, nematodes are living, and so soil samples should be kept cool (do not freeze) during sampling and shipping. To determine the trajectory of nematode damage, it is useful to sample twice a year (Ferris et al. 2004). Nematode sampling can be done at any time of year that the ground is not frozen.

However, fall sampling may be more appropriate since nematode densities will be highest at this time of the season.

The distribution of lesion nematode, and associated potential for replant disease, can be missed with standard soil sample assessment (above) due to the percentage of nematodes inside plant roots (as well as other factors). Mist chamber extraction from root tissue is a more predictable assay. [Oregon State’s Nematode Testing website](#) gives information on how to send a root sample. Plant bioassays also more predictably assess potential damage to new plantings from lesion nematode. Currently, this method is not offered commercially. In brief, a soil sample made up of subsamples from at least fifteen areas in a plot are divided in two where half is pasteurized at 176 to 212 F (80–100 C for 8–12 hours) and apple seedlings grown in pasteurized and unpasteurized soil. Growth is then compared between apple seedlings in pasteurized and unpasteurized soil.

Cultural Controls

New field preparation: It is critical to rip and cultivate soil in order to remove plant roots harboring nematodes, to reduce plant stress to the next planting, and to enhance the efficacy of any fumigation or nematicide applications that may be used.

Resistant Rootstocks

Resistant rootstocks are recommended whenever available (see Table 2).

Green Manures and Fallow

Most plant-parasitic nematodes that are problematic in orchards have a wide host range. Leaving a field fallow may reduce nematode numbers. However, because weeds and other crops such as alfalfa are also hosts, reduction in nematode populations may not be sufficient to reduce crop damage unless a poor host is planted. Consider a green fallow with a poor host (see Table 1).

For dagger nematode, planting orchard ground to a non-host for the virus (such as a grass) can be effective in reducing the concentration or distribution of the virus. Dagger nematodes which feed on the grass cover crop will shed the virus particles present in their esophageal area and will not be able to transfer the virus to fruit trees during later feeding events.

Cultivars of any plant species may differ in their susceptibility to species of nematodes. Similarly, populations of a nematode species may differ in their virulence to plant species or cultivars. Additional research and monitoring may be necessary to determine the suitability of cover crops for management of local nematode populations. We provide a Table of the host status of some cover crop cultivars to populations of nematode species that are considered important in orchards.

Rootstock Selection

Tree architecture and productivity considerations often override nematode concerns in rootstock selection in Washington. Some rootstocks have known tolerance to nematodes.

Biocontrol Options

(** indicates products with an OMRI Label.)

Myrothecium verrucaria

Myrothecium verrucaria (DiTera DF**) is a fungus. Limited greenhouse trials have shown efficacy (Dong et al. 2015). Most commercial products utilize the metabolites produced during the fermentation process in combination with heat-killed mycelium of the fungus. The material may be directly nematicidal, may inhibit nematode egg development, and can reduce the nematode's ability to find the host (Wilson and Jackson 2013).

Paecilomyces lilacinus

Paecilomyces lilacinus (MeloCon WG**) is a fungus which can parasitize nematodes. When spores of this fungus come in contact with nematodes, spores germinate, and fungal hyphae directly parasitize eggs, juveniles, and adult nematodes by excreting enzymes which break down the outer coating of the nematode/egg (chitinase) and direct penetration with an infection peg (appressoria). In greenhouse trials *P. lilacinus* reduced nematodes 30%, and 67% in a field trial (Hashem and Abo-Elyousr 2011; Saha and Khan 2016). The treatment is generally applied in a suspension of water through ground application or drip irrigation. Care must be taken to follow storage requirements of this living product.

Natural Enemies

Plant-parasitic nematodes are food resources for many other organisms, including certain fungi, predaceous and omnivorous nematodes, tardigrades, some collembolan, and others. Unfortunately, many natural enemies are not tolerant of conventional tillage or cultural and fertilization practices common in orchards and vineyards. In particular, many natural enemies are more abundant and active in soils with higher levels of organic carbon. That is because they either use the organic residues as a resource or increase in abundance because they feed on other organisms that utilize the resources. Soil organic matter levels and the natural enemies of nematodes that are supported may be elevated by the use of cover crops or by supplementing organic matter from external sources. Management with organic matter additions requires a consistent program, not just a one-time application. You are attempting to nurture the soil organisms through their interactions. With the use of cover crops, it is important to select species that do not support the plant-parasitic nematode species that you are trying to manage. Examples of some of the principles and ideas that underlie such approaches are provided in Ferris et al. (2012) and Ferris and Tuomisoto (2015).

Table 1. Cover crop host susceptibility.

	Lesion		Ring	Dagger		Root-knot	
	<i>P. penetrans</i>	<i>P. vulnus</i>	<i>C. xenoplax</i>	<i>X. americanum</i>	<i>X. rivesi</i>	<i>M. hapla</i>	<i>M. chitwoodi</i>
Forage Radish	S	U	U	S	U	MR	R
White Mustard	S	U	U	U	U	MR	S
Brown Mustard	S	U	U	U	U	MS	U
Canola	S	U	U	U	U	MS	S
Turnip	S	U	U	U	U	MS	R
Tall Fescue	S	R ^a	U	MR	U	R	U
Wheat	MS	U	R ^d	S	U	R	S
Barley	S	U	MR ^{d?}	S	U	R	R
Arugula	U	U	U	U	U	R	U
Buckwheat	MR	U	U	U	U	R	R
Hairy Vetch	S	U	MS?	U	U	S	U
White Clover	S	U	U	S	U	S	S
Red Clover	S	U	U	S	U	S	S
Crotalaria	S	S	U	R	R	S	U
Lespedeza	S	U	S	U	U	S	U
Red Fescue	S	MR ^a	U	U	U	S?	S
Pearl Millet	R ^b	U	U	U	U	S?	U
Corn	S	U	S?	S	U	MR	S
Rye	S	U	MR ^d	S?	U	S?	MS
Oats	S	U	MS ^{d?}	S	U	S?	S
Annual Ryegrass	S	U	MS	U	U	S?	U
Field Peas	S	S	U	U	U	S?	S
Perennial Ryegrass	S	MR ^a	MS	U	U	U	U
Sudan Grass	S	U	S?	R	U	R ^c	R
Triticale	U	U	MR	U	U	U	U
Teff	U	U	U	U	U	U	U
Cowpea	S	S	U	S	U	MR	R

S = susceptible, R = resistance, MR = medium resistance, MS = medium susceptible, U = undetermined, ? = conflicting literature. Nemabase (Ferris 2017). ^a(Westerdahl 2006), ^b(Dauphinais et al. 2005), ^d(Nyczepir and Bertrand 1990), ^c(Mojtahedi et al. 1993).

Table 2. Nematode resistance in Prunus rootstocks.

	Lesion		Ring	Dagger		Root-knot		
	<i>P. penetrans</i>	<i>P. vulnus</i>	<i>M. xenoplax</i>	<i>X. americanum</i>	<i>X. rivesi</i>	<i>M. hapla</i>	<i>M. chitwoodi</i>	<i>M. incognita</i>
Cadaman		S	S					R
Citation		S						R
Colt	S		S	S				
Halford								
Ishtara								
Jaspi								
Julior								
Krymsk 86		S						S
Lovell		S	MS					S
Mahaleb	S		S	S				
Marianna 26-23		S	S					
Marianna 26-24		S	S	S		R		
Marianna GF 8-1		S						R
Maxma 14								
Mazzard	MR	S	S	S				
MXM-14	S	S	S	S				
MXM-60	S		S	S				
MXM-97	S		S					
Myrobalan								
Myrobalan 29C		S	S	S		R		R
Nemaguard	S	S	S	S		R		R
Paramount		MS						S
Pumiselect		S						R
Torinel								

S = Susceptible, R = Resistance, MS = Medium Susceptible, MR = Medium Resistance. From Nemabase, updated Ferris 2017. Includes rootstocks grown in Washington. For a complete Table visit Nemabase.

Chemical Nematicides

K-Pam HL

K-Pam HL (metam-potassium) is registered for sprinkler application on orchard replant sites in Washington.

Luna Privilege

Luna Privilege (fluopyram) is registered for chemigation in stone fruit and pome fruit orchards in Washington.

Fumigants

A mixture of 1,3-dichloropropene (1,3-D) and chloropicrin (CP): Telone C35 (63% 1,3-dichloropropene and 35% chloropicrin) is one of the most widely used pre-plant fumigants. It provides acceptable control of nematodes and soil borne pathogens. Virtually impermeable films (VIF) and totally impermeable films (TIF) improve retention of fumigant volatiles in the soil resulting in improved efficacy and reduced chemical emissions (Cabrera et al. 2015, Gao et al. 2016).

Note: Chemical nematicides and fumigants may have harmful effects on the organisms of the soil food web that contribute to biological regulation of pest nematode species.

Washington Nematode Testing Labs

[AgNema](#), Prosser, WA

[Soil Test](#), Moses Lake, WA, (509) 765-1622

[A&L Western Laboratories](#)

[Oregon State University Plant Clinic](#), (541) 737-3472

WSU online database for [Analytical Testing Services](#)

Additional Reading

Ferris, H. 1999 (Updated 2018). [The Nematode Plant Expert Information System](#). Nemaplex. University of California.

Use pesticides with care. Apply them only to plants, animals, or sites listed on the labels. When mixing and applying pesticides, follow all label precautions to protect yourself and others around you. It is a violation of the law to disregard label directions. If pesticides are spilled on skin or clothing, remove clothing and wash skin thoroughly. Store pesticides in their original containers and keep them out of the reach of children, pets, and livestock.

YOU ARE REQUIRED BY LAW TO FOLLOW THE LABEL. It is a legal document. Always read the label before using any pesticide. You, the grower, are responsible for safe pesticide use. Trade (brand) names are provided for your reference only. No discrimination is intended, and other pesticides with the same active ingredient may be suitable. No endorsement is implied.

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